



# A new genus of metalmark moths (Lepidoptera, Choreutidae) with Afrotropical and Australasian distribution

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#### **Abstract**

Niveas Rota, new genus, and its two new species, N. agassizi Rota, new species, and N. kone Rota, new species, are described and illustrated. Niveas is assigned to the subfamily Choreutinae based on morphological and molecular data. Niveas agassizi is currently known only from Kenya and only from female specimens. Niveas kone has been found on the Solomon Islands and in Papua New Guinea (PNG). In PNG, larvae of this species have been reared from several species of Ficus (Moraceae). The two species are superficially quite dissimilar from each other. However, they share features in wing pattern and venation, as well as female genitalia, and the molecular data strongly support the monophyly of Niveas.

#### Keywords

Alpha taxonomy, DNA barcoding, Ficus spp., Kenya, Niveas agassizi, Niveas kone, Papua New Guinea, Solomon Islands, phylogenetics

## Introduction

Choreutidae, commonly known as metalmark moths, are a family of micro-moths with a worldwide distribution. The family is most species-rich in the tropics, and, as is the case for numerous other small tropical invertebrates, much of its richness is still unknown to science (unpublished data). Currently, 406 species of choreutids are described (Nieukerken et al. 2011).

Choreutids are medium-sized micro-moths with wingspans ranging from about one to two centimeters, often with bright colors and iridescent markings on their wings (Diakonoff 1986). They are diurnal with only some species attracted to lights at night (personal observation), making them a fairly rare group in museum collections. In our experience, large-scale rearing projects result in finding more species of choreutids than employing light traps.

Through exactly such efforts over the past 20 years in Papua New Guinea (PNG), the Binatang Research Center (BRC), with a large international group of collaborators focusing on the ecology of herbivorous insects and their host plants (Miller et al. 2003; Craft et al. 2010; Novotny et al. 2010; Hrcek et al. 2011; Hrcek et al. 2013; Miller et al. 2013), the number of known species of choreutids and our knowledge of their biology have greatly increased. One of the many new species of choreutids found in PNG during this project is sufficiently different from all described species that it requires a new genus.

Coincidentally, through separate collecting efforts by David Agassiz in Africa, a related species was discovered in Kenya. Herein these two species, as well as the genus to which they belong, are described and illustrated, and the phylogenetic position of the new genus within the family is discussed.

The shared presence of the terminal black band with white spots in the forewing (arrows in Figs 1, 3) was the first indication that *N. kone* Rota, sp. n. and *N. agassizi* Rota, sp. n. might be related. Initially this relationship seemed unlikely because of the disjunct geographical distribution of the two (*N. kone* being distributed in the Australasian Region and *N. agassizi* in the Afrotropical Region) and because their DNA barcodes did not suggest a close relationship. However, once the similarities in wing venation and female genitalia were noticed, and we included nuclear genes in the analysis with a more extensive choreutid molecular dataset, the results strongly supported the close relationship between *N. kone* and *N. agassizi*.

### **Methods**

All material examined is listed in Table 1. Layered photographs of specimens and slides were taken using an Olympus SZX16 microscope with motorized focus drive attached to an Olympus E520 digital camera. The photographs were then combined by using the programs Deep Focus 3.1 and Quick Photo Camera 2.3. The wing venation drawing was made digitally in Adobe Illustrator CS3 overlaid on top of a slide photograph.

**Table 1.** Material examined.

Species	Type	Country	Province Locality	Locality	Date	Collector	ID number	Host plant	Slide number	GenBank
N. kone	Paratype	PNG	Madang	Baitabag Vill.	04/09/95	BRC	USNM ENT 730507	Ficus nodosa		
N. kone	Paratype	PNG	Madang	Baitabag Vill.	08/30/95	BRC	USNM ENT 730558	Ficus nodosa		
N. kone	Paratype	PNG	Madang	Baitabag Vill.	96/30/80	BRC	<b>USNM ENT 730572</b>	Ficus nodosa		HQ946542
N. kone	Paratype	PNG	Madang	Baitabag Vill.	06/16/95	BRC	<b>USNM ENT 730508</b>	Ficus variegata		
N. kone	Paratype	PNG	Madang	Baitabag Vill.	03/19/96	BRC	USNM ENT 730513	Ficus variegata		HQ946551
N. kone	Paratype	PNG	Madang	Baitabag Vill.	04/09/95	BRC	<b>USNM ENT 730529</b>	Ficus variegata		KF714836
N. kone	Paratype	PNG	Madang	Baitabag Vill.	03/19/96	BRC	<b>USNM ENT 730543</b>	Ficus variegata		
N. kone	Paratype	PNG	Madang	Baitabag Vill.	03/19/96	BRC	USNM ENT 730551	Ficus variegata		
N. kone	Paratype	PNG	Madang	Kamba (Mis)	10/20/95	BRC	<b>USNM ENT 730576</b>	Ficus variegata		HQ946555
N. kone	Paratype	PNG		Malapau (Riwo)	03/20/95	BRC	USNM ENT 730498	Ficus variegata		HQ946554
N. kone	Paratype	PNG	Madang	Malapau (Riwo)	03/20/95	BRC	<b>USNM ENT 730519</b>	Ficus variegata		HQ946553
N. kone	Paratype	PNG	Madang		03/20/95	BRC	USNM ENT 730535	Ficus variegata		HQ946552
N. kone	Paratype	PNG	Madang	Mililat (Riwo)	05/22/95	BRC	USNM ENT 730604	Ficus nodosa		HQ946544
N. kone	Paratype	PNG	Madang	Mis Vill.	03/20/96	BRC	<b>USNM ENT 730528</b>	Ficus nodosa		HQ946543
N. kone	Paratype	PNG	Madang	Ohu Vill.	04/09/95	BRC	<b>USNM ENT 730560</b>	Ficus botryocarpa		HQ946538
N. kone	Paratype	PNG	Madang	Ohu Vill.	05/09/95	BRC	<b>USNM ENT 730602</b>	Ficus botryocarpa		HQ946539
N. kone	Paratype	PNG	Madang	Ohu Vill.	12/01/96	BRC	<b>USNM ENT 730542</b>	Ficus phaeosyce		KF714835
N. kone	Paratype	PNG	Madang	Ohu Vill.	12/02/94	BRC	<b>USNM ENT 730502</b>	Ficus pungens		HQ946546
N. kone	Paratype	PNG	Madang	Ohu Vill.	12/09/95	BRC	<b>USNM ENT 730518</b>	Ficus variegata	female	HQ946549
									genitalia 92352	
N. kone	Paratype	PNG	Madang	Ohu Vill.	03/16/95	BRC	<b>USNM ENT 730509</b>	Ficus variegata	male	HQ946550
									genitalia 92355	
N. kone	Paratype	PNG	Madang	Ohu Vill.	03/16/95	BRC	<b>USNM ENT 730492</b>	Ficus variegata		
N. kone	Paratype	PNG	Madang	Ohu Vill.	03/25/96	BRC	<b>USNM ENT 730493</b>	Ficus variegata		
N. kone	Paratype	PNG	Madang	Ohu Vill.	96/60/50	BRC	USNM ENT 730500	Ficus variegata		

Species	Type	Country	Province Locality	Locality	Date	Collector	ID number	Host plant	Slide number	GenBank
N. kone	Paratype	PNG	Madang	Ohu Vill.	03/22/95	BRC	<b>USNM ENT 730504</b>	Ficus variegata		
N. kone	Paratype	PNG	Madang	Ohu Vill.	12/13/94	BRC	<b>USNM ENT 730510</b>	Ficus variegata		
N. kone	Holotype	PNG	Madang	Ohu Vill.	03/13/95	BRC	<b>USNM ENT 730516</b>	Ficus variegata		HQ946548
N. kone	Paratype	PNG	Madang	Ohu Vill.	56/60/80	BRC	<b>USNM ENT 730517</b>	Ficus variegata		
N. kone	Paratype	PNG	Madang	Ohu Vill.	03/16/95	BRC	USNM ENT 730520	Ficus variegata		
N. kone	Paratype	PNG	Madang	Ohu Vill.	05/26/95	BRC	USNM ENT 730521	Ficus variegata		
N. kone	Paratype	PNG	Madang	Ohu Vill.	05/09/95	BRC	<b>USNM ENT 730522</b>	Ficus variegata		
N. kone	Paratype	PNG	Madang	Ohu Vill.	03/29/95	BRC	<b>USNM ENT 730523</b>	Ficus variegata		
N. kone	Paratype	PNG	Madang	Ohu Vill.	06/16/95	BRC	USNM ENT 730524	Ficus variegata		
N. kone	Paratype	PNG	Madang	Ohu Vill.	05/11/96	BRC	<b>USNM ENT 730525</b>	Ficus variegata		
N. kone	Paratype	PNG	Madang	Ohu Vill.	06/27/95	BRC	<b>USNM ENT 730526</b>	Ficus variegata		
N. kone	Paratype	PNG	Madang	Ohu Vill.	06/16/95	BRC	<b>USNM ENT 730531</b>	Ficus variegata		
N. kone	Paratype	PNG	Madang	Ohu Vill.	12/13/94	BRC	USNM ENT 730533	Ficus variegata		
N. kone	Paratype	PNG	Madang	Ohu Vill.	96/60/50	BRC	<b>USNM ENT 730553</b>	Ficus variegata		
N. kone	Paratype	PNG	Madang	Ohu Vill.	12/09/95	BRC	<b>USNM ENT 730564</b>	Ficus variegata		
N. kone	Paratype	PNG	Madang	Ohu Vill.	05/09/95	BRC	<b>USNM ENT 730588</b>	Ficus variegata		
N. kone	Paratype	PNG	Madang	Ohu Vill.	05/09/95	BRC	<b>USNM ENT 730595</b>	Ficus variegata		
N. kone	Paratype	PNG	Madang	Ohu Vill.	09/10/95	BRC	<b>USNM ENT 730565</b>	Ficus wassa		HQ946545
N. kone	Paratype	PNG	Madang	Pau Vill.	12/13/95	BRC	<b>USNM ENT 730515</b>	Ficus variegata		KF714837
N. kone	Paratype	PNG	Madang	Pau Vill.	12/13/95	BRC	<b>USNM ENT 730547</b>	Ficus variegata		KF714833
N. kone	Paratype	PNG	Madang	Reinduk	03/28/95	BRC	<b>USNM ENT 730527</b>	Ficus variegata		KF714834
N. kone	Paratype	PNG	Madang	Tab Is	01/31/95	BRC	<b>USNM ENT 730506</b>	Ficus nodosa		
N. kone	Paratype	PNG	Madang	Tab Is	01/31/95	BRC	<b>USNM ENT 730514</b>	Ficus nodosa		KF714832
N. kone	Paratype	PNG	Madang	Tab Is	01/31/95	BRC	<b>USNM ENT 730532</b>	Ficus nodosa		HQ946540
N. kone	Paratype	PNG	Madang	Tab Is	01/31/95	BRC	<b>USNM ENT 730538</b>	Ficus nodosa		HQ946541
N. kone	Paratype	PNG	Madang	Wanang Vill.	07/31/07	BRC	USNM ENT 660733	Ficus variegata		
N. kone	Paratype	PNG	Madang	Wanang Vill.	11/05/07	BRC	USNM ENT 660794	Ficus variegata		
N. kone	Paratype	PNG	Madang	Wanang Vill.	02/21/06	BRC	<b>USNM ENT 660722</b>	unknown		HQ946547

Species	Type	Country	Province	Locality	Date	Collector	ID number	Host plant	Slide number	GenBank
N. kone	Paratype	Solomon Is.		Guadalca- Roroni, 35 km nal E of Honiara; 10 m	05/13/64	R. Straat- man	unassigned	unknown	wing 137601; female genitalia 137600	
N. kone	Paratype	Solomon Is.	Guadalca- nal	Guadalca- Roroni, 35 km nal E of Honiara; 10 m	05/13/64	R. Straat- man	unassigned	unknown		
N. kone	Paratype	Solomon Is.		Guadalca- Nini Ck., 35 km 08/05/64 nal SE of Honiara	08/05/64	R. Straat- man	unassigned	unknown		
N. agassizi	Paratype	Kenya	County of Kwale	Mwabungu	08/19/00	David Agassiz	USNM ENT 730794	unknown		HQ946715
N. agassizi	Holotype	Kenya	County of Kwale	Mwabungu	08/19/00	David Agassiz	USNM ENT 730793	unknown		HQ946716
N. agassizi	Paratype	Kenya	County of Kwale	Mwabungu	08/19/00	David Agassiz	unassigned	unknown	female genitalia 137597	
N. agassizi	Paratype	Kenya	County of Kwale	Mwabungu	08/19/00	David Agassiz	unassigned	ипкпошп	female genitalia JR2013- 02	
N. agassizi	Paratype	Kenya	County of Kwale	Mwabungu	08/19/00	David Agassiz	unassigned	ипкпошп	wing JR2013- 03	
N. agassizi	Paratype	Kenya	County of Kwale	Mwabungu	08/20/00	David Agassiz	unassigned	unknown	female genitalia JR2013- 01	

All images were improved in Adobe Photoshop CS3. Genitalic dissections and terminology follow Rota (2008b).

Field sampling and rearing protocols for the PNG material are detailed in Miller et al. (2003; 2013), Craft et al. (2010), and Novotny et al. (2010). The Plant List website (2010) was used for host plant names. Latitude, longitude, and altitude data for the collecting localities is in Table 2.

The molecular phylogeny dataset included three outgroups and 40 species of ingroup taxa, including two individuals each of Niveas kone and N. agassizi totaling 45 terminal units. We analyzed data from eight genes: COI (mitochondrial), CAD, EF1a, GAPDH, IDH, MDH, RpS5, and wingless (all nuclear) (Wahlberg and Wheat 2008). The final alignment was 6187 base pairs long. Molecular sequences for all taxa except N. kone and N. agassizi are from Rota (2011) and Rota and Wahlberg (2012), and their GenBank accession numbers can be found there. For the specimens of *N. kone* (660733) and N. agassizi (Ch\_JR44\_1), DNA extraction was done from whole abdomens, which were later used for dissection of genitalia. Because the DNA amplification methods described by Wahlberg and Wheat (2008) did not work for obtaining sequences of nuclear genes from these specimens, suggesting that their DNA was too degraded for the standard approach, we used newly-designed primers (Niklas Wahlberg, unpublished) (Table 3) to amplify short fragments of the nuclear genes (see Table 4 for total number of base pairs for each gene fragment amplified and the GenBank accession numbers for fragments longer than 200 base pairs). For sequence storage and manipulation we used the VoSeq application (Peña and Malm 2012). The nexus file with the alignment is available from the Figshare Digital Repository: doi: 10.6084/m9.figshare.811841

Both maximum likelihood (ML) and Bayesian phylogenetic analyses were performed. ML analysis of unpartitioned data was conducted using RAxML blackbox available online (Stamatakis et al. 2008) with the GTR+G model and 100 bootstraps. Bayesian analysis of data partitioned using the program TIGER (Cummins and McInerney 2011) as described in Rota and Wahlberg (2012) was carried out in MrBayes v. 3.2 (Ronquist et al. 2012) for 10 million generations with one cold and three heated chains, sampling trees every 1000 generations. The analyses were run on the freely available Bioportal server (University of Oslo, Norway). The convergence was assessed

Table 2	<b>2.</b> Loca	lity info	ormation.
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Locality	m.a.s.l.	latitude	longitude	
Baitabag village & Kau Wildlife Area, near Madang, Madang Province, PNG	50	S5°08'	E145°46'	_
Mis, Madang Province, PNG	50	S5°11'	E145°47'	_
Ohu Conservation Area, Ohu village near Gum river, Madang Province, PNG	100	S5°13'	E145°41'	
Pau, Madang Province, PNG	0	S5°08'	E145°46'	_
Reinduk, Madang Province, PNG	225	S5°39'	E145°24'	_
Riwo, Madang Province, PNG	0	S5°09'	E145°48'	_
Tab Island, Madang Province, PNG	0	S5°10.6'	E145°52.6'	_
Wanang village, Madang Province, PNG	115	S5°13.9'	E145°10.9'	_
Mwabungu, County of Kwale, Kenya	0	S4°20.3'	E39°37'	_

by examining plots of log likelihoods and all model parameters using Tracer v.1.5 (Rambaut and Drummond 2007), as well as potential scale reduction factors and split frequencies, both reported by MrBayes. Branch support is expressed as Bayesian posterior probability (PP) and maximum likelihood bootstraps (ML BS).

DNA barcode sequences (COI) for *Niveas kone* (24 specimens) and *Niveas agassizi* (2 specimens) were obtained at the Biodiversity Institute of Ontario, University of

Table 3. Primers.

COI-1F	GGTCAACAAATCATAAAGATATTGG
COI-1R	GGWGCYCCTARTATTAAAGGWAYTA
EF-1F	CACATYAACATTGTCGTSATYGG
EF-1R	TRSCGGTYTCGAACTTCCA
EF-2F	GAGCGTGARCGTGGTAT
EF-2R	RGCTTCGAACTCACCRGTA
EF-3F	TCAAGAACATGATCACYGG
EF-3R	GARGAYACTTCCTTGA
EF-7F	CAAYGTTGGTTTCAACGT
EF-8R	ACAGCVACKGTYTGYCTCATRTC
GAPDH-1F	AARGCTGGRGCTGAATATGT
GAPDH-1R	AAGTTGTCATGGATRACCTT
GAPDH-2F	GTCATCTCYAATGCYTCYTG
GAPDH-2R	TAACTTTGCCRACAGCYTT
GAPDH-3F	GTGCCCARCARAACATCAT
GAPDH-3R	TCAGCGGCTTCCTTRACCT
IDH-1F	GGWGAYGARATGACNAGRATHATHTGG
IDH-1R	GGACTCTTCCACATTTTYTT
MDH-1F	GAYATNGCNCCNATGATGGGNGT
MDH-1R	TCYTTRCGRGCAACYTTRTC
RPS5-1F	ATGGCNGARGARAAYTGGAAYGA
RPS5-1R	TTGTGWGCRTACCTRCCRGC

**Table 4.** GenBank accession numbers and the number of base pairs for each gene fragment.

	N. agassizi (730793)	N. agassizi (Ch_JR44_1)	N. kone (730509)	N. kone (660733)
COI	HQ946716	-	HQ946550	KF646130
COI	609 bp	176 bp	658 bp	610 bp
EF1a	-	KF646128, KF646129	-	KF646131, KF646132
	-	550 bp	-	706 bp
GAPDH	-	-	-	KF646133
	-	136 bp	-	430 bp
IDH	-	135 bp	-	-
MDH	-	190 bp	-	-
RpS5	-	155 bp	-	108 bp

Guelph, using their standard methodology (Craft et al. 2010; Hrcek et al. 2011; Wilson 2012). They are deposited in GenBank as accessions listed in Table 1, and their full data including images are in the Barcode of Life Database (http://www.boldsystems.org; see Ratnasingham and Hebert 2007; 2013). These sequences were also analyzed with MrBayes v. 3.2 (unpartitioned dataset, 2 million generations).

### Results

## **Taxonomy**

## Niveas Rota, gen. n.

http://zoobank.org/F352952E-0F21-464F-BD1E-278C9A0679C1 http://species-id.net/wiki/Niveas Figs 1–9

## Type species. Niveas kone.

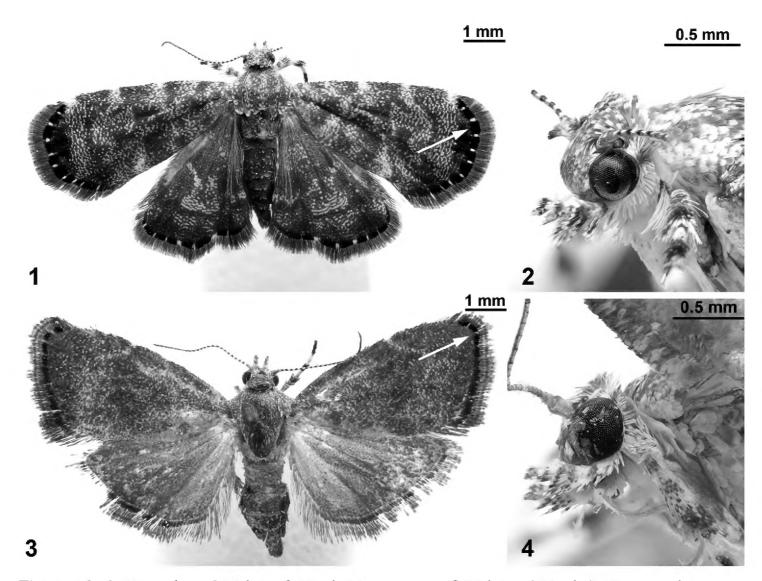
**Material examined.** See Table 1.

Distribution. Kenya, Papua New Guinea, Solomon Islands.

**Diagnosis.** Niveas can be easily distinguished from most genera of choreutids by the wing pattern (Figs 1, 3). Superficially, species of Niveas are similar to some species of Anthophila and Choreutis, but there is no known species in either of the latter two genera with a black terminal band enclosing white spots in the forewing as in N. agassizi and N. kone. (Figs 1, 3). Forewing venation with only four radial branches or with R<sub>4</sub> and R<sub>5</sub> fused in the basal half is also diagnostic for the genus. Female genitalia with paired concave sclerotizations on A7 sternite are also unique to Niveas.

**Description.** Head. Labial palpi with projecting ventral scale tufts (Figs 2, 4). Wings. Forewing veins R four-branched in N. kone (Fig. 5), five-branched in N. agassizi (Fig. 6), with R<sub>4</sub> an R<sub>5</sub> fused in basal 3/5; CuP present at termen for 1/3 to 1/5 wing length, extending as fold further towards base. Hindwing ten-veined, with M<sub>2</sub> in close proximity to the basally fused M<sub>3</sub> and CuA<sub>1</sub> (N. agassizi) or nine-veined, apparently with M<sub>3</sub> and CuA<sub>1</sub> completely fused into a single vein (Figs 5, 6). Male genitalia. Tegumen rounded on top, tuba analis extending beyond tegumen; vinculum as inverted trapezoid ventrally emarginate; valva with costal margin straight, ventral margin rounded, ending with a horn-like projection; phallus twice as long as valva (Fig. 7). Female genitalia. Apophyses anteriores slightly longer than posteriores; ostium bursae on A7 with a more or less strongly sclerotized antrum; ductus bursae straight, not coiled, with strong lateral sclerotizations; corpus bursae as a single sac (N. agassizi) or divided into two sacs (N. kone) with one or more signa. A7 sternite with paired, somewhat rounded, concave sclerotizations proximally, clearly visible in N. kone (Fig. 8), and slightly less so in N. agassizi (Fig. 9).

Host plants. Genus Ficus (Moraceae).



**Figures 1–4.** *Niveas kone*: **I** Habitus **2** Head. *Niveas agassizi*: **3** Habitus **4** Head. (In Figs 1 and 3 arrows point at the terminal black band enclosing white spots.)

**Etymology.** The generic name is derived from Latin *niveum*, meaning snowy, in reference to speckles of white-tipped scales in the wings of the type species; it is not treated as a Latin word and is feminine in gender.

## Niveas kone Rota, sp. n.

http://zoobank.org/9EA367B0-6B92-48FA-8075-D8D0D0BFA566 http://species-id.net/wiki/Niveas\_kone Figs 1, 2, 5, 7, 8

#### **Material examined.** See Table 1.

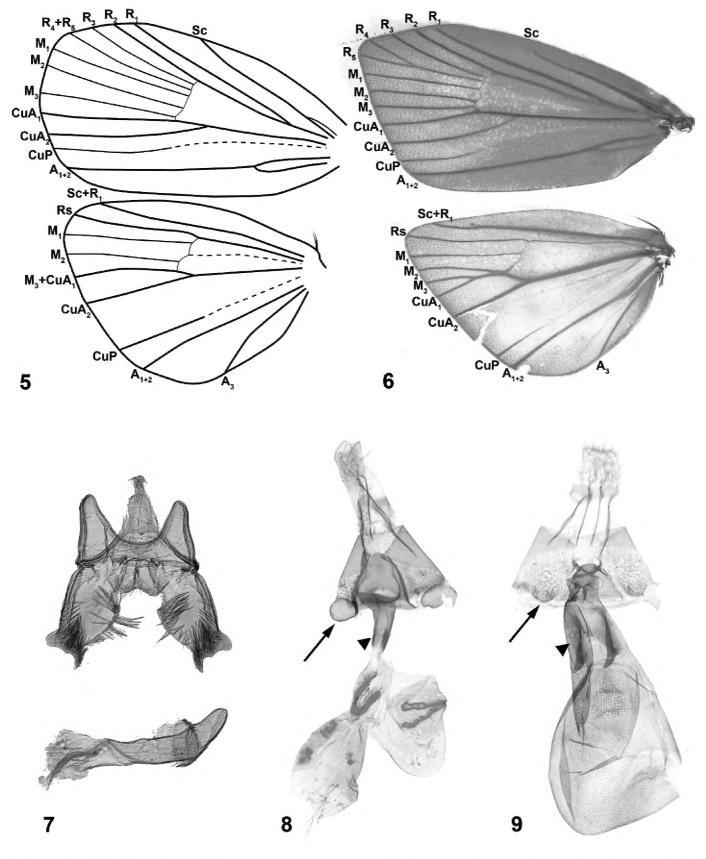
**Material deposited.** The holotype and most paratypes will be retained at USNM, with paratypes distributed to PNG National Agriculture Research Institute (Port Moresby), BMNH, Bishop Museum, Naturalis (Leiden), and CSIRO (Canberra).

**Distribution.** Papua New Guinea, Solomon Islands.

**Diagnosis.** N. kone can be separated from all other known choreutids based on its wing pattern (Fig. 1). Superficially, it is similar to a few species of *Brenthia* Clemens, 1860 and *Litobrenthia* Diakonoff, 1978 owing to its background color, but it lacks iridescent

spots along forewing termen, which are always present in those two genera. Both male and female genitalia are very distinct from those of other choreutids (Figs 7, 8).

**Description.** Head. Fig. 2. Wings. Fore- and hindwing with brown background color, speckled with white-tipped scales in an irregular pattern; a distinct black band along termen of both wings within which are more or less equidistant white spots (Fig. 1). Male genitalia. As for the genus (Fig. 7). Female genitalia. Corpus bursae split



**Figures 5–9.** *Niveas kone*: **5** Wing venation **7** Male genitalia **8** Female genitalia. *Niveas agassizi*: **6** Wing venation **9** Female genitalia. (In Figs 8 and 9 arrows point at the A7 sternite sclerotizations, and triangles point at the lateral sclerotizations on the ductus bursae.)

into two sacs; one sac with a V-shaped signum, the other with two round signa (Fig. 8). *Immature stages.* Fig. 12. See a brief note in text.

**Host plants.** Ficus botryocarpa Miq., F. nodosa Teijsm. & Binn., F. phaeosyce K. Schum. & Lauterb., F. pungens Reinw. ex Blume, F. variegata Blume, and F. wassa Roxb. (Moraceae).

**Etymology.** The species is named after the Finnish Kone Foundation (Koneen Säätiö) in appreciation of their funding of this work. The name is a noun in apposition.

## Niveas agassizi Rota, sp. n.

http://zoobank.org/7F08322B-C0D2-450C-9DFF-ED9E4FEA5892 http://species-id.net/wiki/Niveas\_agassizi Figs 3, 4, 6, 9

#### **Material examined.** See Table 1.

**Material deposited.** The holotype will be deposited in National Museums of Kenya (Nairobi) (NMK), with paratypes to USNM, BMNH and NMK.

**Distribution.** Kenya.

**Diagnosis.** *N. agassizi* can be separated from other known choreutids by the wing pattern (Fig. 3). It is superficially similar to some species of *Choreutis*, but the latter usually have forewings with apparent patterning, and this is absent in *N. agassizi*. Female genitalia are very distinct from those of other choreutids (Fig. 9).

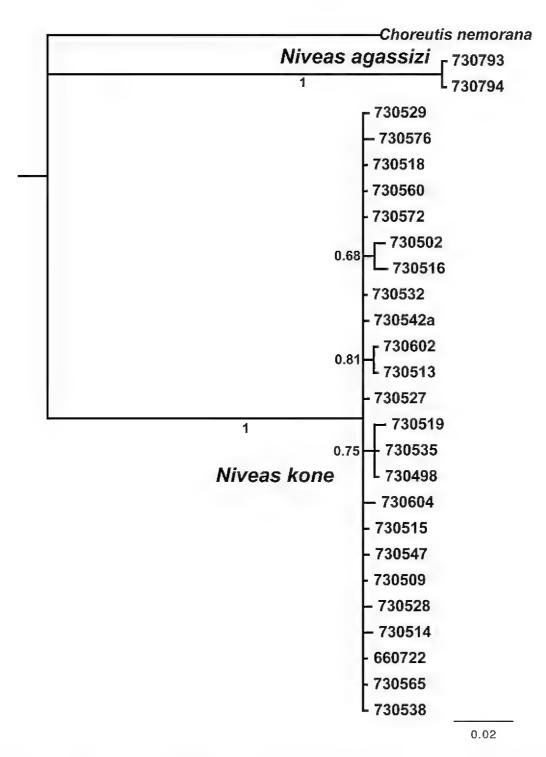
**Description.** Male unknown. *Head.* Fig. 4. *Wings.* Forewing bronze-brown with speckled white-tipped scales over most of its surface; distinct dark brown to black band along termen with two small white spots at apex; hindwing light brown (Fig. 3). *Male genitalia.* Unknown. *Female genitalia.* Ductus bursae short and wide, opening into large corpus bursae, with one oval signum (Fig. 9). *Immature stages.* Unknown.

Host plants. Unknown.

**Etymology.** This species is named after David Agassiz, who collected all the known specimens and made many significant contributions to our knowledge of African micro-moths. The name is a noun in the genitive case.

#### Remarks

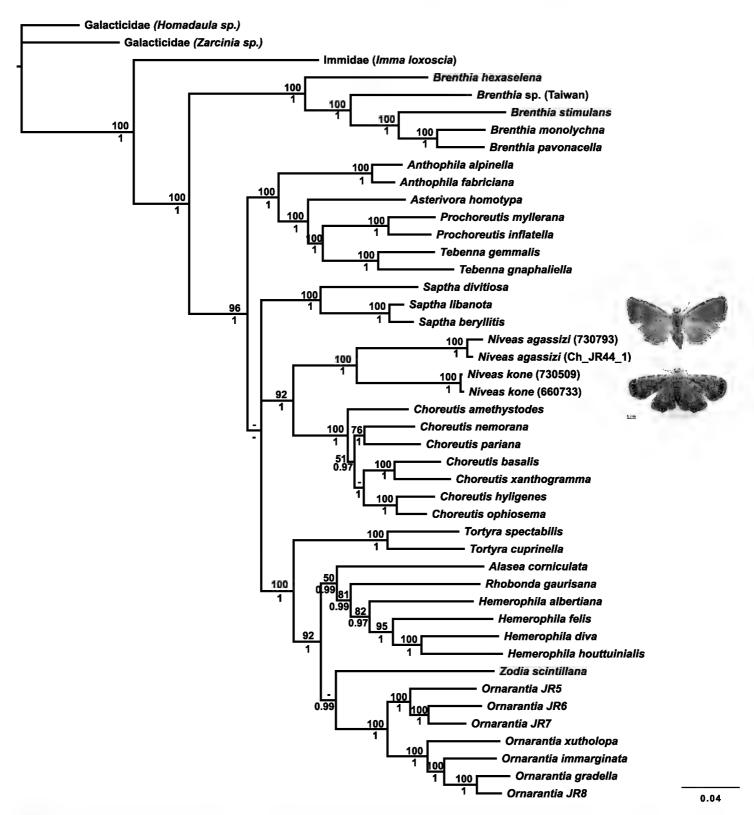
We obtained 19 full-length barcodes of *Niveas kone*, as well as 5 shorter fragments. These form cluster AAB7478 in the Barcode of Life Database (accessed 29 August 2013), and using the RESL algorithm as implemented there (Ratnasingham and Hebert 2013), the maximum distance between the COI sequences for members of the species is 0.65%, whereas the distance to the nearest cluster (*N. agassizi*) is 9.22%. In a Bayesian analysis of the COI sequences, all *N. kone* and all *N. agassizi* specimens grouped together with the other members of their species with very high branch support (PP=1) (Fig. 10).



**Figure 10.** DNA barcode tree from a Bayesian analysis showing low divergence within species and high between species of *Niveas*. Numbers below or next to branches are Bayesian posterior probabilities. Specimen ID numbers are used as labels for the terminal branches.

The placement of *Niveas* in the choreutid generic phylogeny is very strongly supported. *Niveas* clearly belongs within the subfamily Choreutinae (PP=1; ML BS=96), and it appears to be the sister group of *Choreutis* (PP=1.00; ML BS=92) (Fig. 11).

Further comments on the biology of *Niveas kone*: Over the years, BRC field teams have encountered larvae identified as *Niveas kone* (as project morphospecies TORT015) 118 times, of which 62 were reared to adults, usually on *Ficus nodosa* and *Ficus variegata*, but also on four other species of *Ficus* (see full host plant list under *N. kone* description). Larvae have been found in all months except April and November, and are described by BRC staff as being green-clear-whitish in color, with short white hairs, and one spot on the side of the head (Fig. 12). Larvae of *N. kone* share the presence of short hairs with other Choreutinae (Rota 2005), which is unlike Brenthiinae



**Figure 11.** Phylogenetic tree from a Bayesian analysis showing the position of *Niveas* in relation to other choreutid genera. Maximum likelihood (ML) bootstraps are shown above branches, and Bayesian posterior probabilities (PP) are below branches; dashes represent ML bootstraps<50 and PP<0.95.

larvae, which possess very long hairs (Rota 2008a). Project field notes indicate that the shelters are distinct from other local Choreutidae in having strong white webbing. BRC has encountered them most commonly in the lowland coastal areas around Madang (city), but also in the coastal mountains behind Madang (up to about 100 m elevation), and at Wanang in the Ramu River Basin (115 m). The species has been recorded in publications (e.g., supplement to Novotny et al. 2010) and online databases as TORT015, misidentified as *Brenthia* sp. Based on locality information provided by Taylor and Maffi (1978: 185, 212), the Solomon Islands specimens are from lowland and foothill localities near Honiara, Guadalcanal; they were collected in light traps.

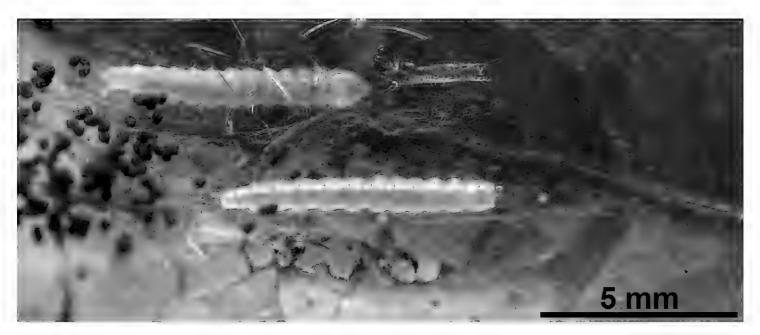


Figure 12. A photograph of the Niveas kone larvae made in the field.

Taxon descriptions are also organized in tabular format for ease of comparison (see Appendix).

## **Discussion**

The two species of *Niveas* described herein are superficially quite different, but upon closer examination it becomes apparent that they share a number of morphological features. We consider the following as potential autapomorphies of *Niveas*: fusion or reduction in R veins in the forewing (Figs 5, 6); presence of round, concave sclerotizations on the A7 sternite in females (arrows in Figs 8, 9); strong lateral sclerotizations at the base of the ductus bursae (triangles in Figs 8, 9); and the presence of a terminal black band with white spots in the forewing (arrows in Figs 1, 3). In all other Choreutinae genera there are five fully-separated radial veins in the forewing; the A7 sternite in the female, as well as the base of the ductus bursae, are evenly sclerotized; and if present, a black terminal band in the forewing lacks white spots.

The split between *N. kone* and *N. agassizi* has presumably happened a long time ago based on the large COI divergence between them and the length of branches in the phylogenetic analysis including the nuclear genes. We considered assigning each species to its own monotypic genus because of their different external appearance, as well as some of the differences in venation and some aspects of female genitalia. It is unfortunate that *N. agassizi* is known from females only as perhaps the morphology of the male genitalia would help clarify the status of this species. However, we believe that *N. kone and N. agassizi* being each other's closest relatives among the currently known species of choreutids is best conveyed by assigning them to a single genus and therefore we opted for this more conservative approach. It is conceivable that other species of *Niveas* that might bridge this gap in both genetic and morphological variation will be discovered in the future. On the other hand, it is also possible that a new genus will need to be erected to accommodate *N. agassizi* and its currently unknown relatives.

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#### References

- Craft KJ, Pauls SU, Darrow K, Miller SE, Hebert PDN, Helgen LE, Novotny V, Weiblen GD (2010) Population genetics of ecological communities with DNA barcodes: An example from New Guinea Lepidoptera. Proceedings of the National Academy of Sciences of the United States of America 107: 5041–5046. doi: 10.1073/pnas.0913084107
- Cummins CA, McInerney JO (2011) A Method for Inferring the Rate of Evolution of Homologous Characters that Can Potentially Improve Phylogenetic Inference, Resolve Deep Divergence and Correct Systematic Biases. Systematic Biology 60: 833–844. doi: 10.1093/sysbio/syr064
- Diakonoff A (1986) Glyphipterygidae *auctorum sensu lato*: (Glyphiterygidae *sensu* Meyrick, 1913); Tortricidae: Hilarographini, Choreutidae, Brachodidae (partim), Immidae and Glyphipterygidae. G. Braun, Druckerei und Verlage, Karlsruhe, Plates Volume (175 pls.) + 436 pp.
- Hrcek J, Miller SE, Quicke DLJ, Smith MA (2011) Molecular detection of trophic links in a complex insect host-parasitoid food web. Molecular Ecology Resources 11: 786–794. doi: 10.1111/j.1755-0998.2011.03016.x

- Hrcek J, Miller SE, Whitfield JB, Shima H, Novotny V (2013) Parasitism rate, parasitoid community composition and host specificity on exposed and semi-concealed caterpillars from a tropical rainforest. Oecologia 173: 521–532. doi: 10.1007/s00442-013-2619-6
- The Plant List (2010) Version 1. http://www.theplantlist.org/ [accessed August 5.2013]
- Miller SE, Hrcek J, Novotny V, Weiblen GD, Hebert PDN (2013) DNA barcodes of caterpillars (Lepidoptera) from Papua New Guinea. Proceedings of the Entomological Society of Washington 115: 107–109. doi: 10.4289/0013-8797.115.1.107
- Miller SE, Novotny V, Basset Y (2003) Studies on New Guinea moths. 1. Introduction (Lepidoptera). Proceedings of the Entomological Society of Washington 105: 1034–1042.
- Nieukerken EJ van, Kaila L, Kitching IJ, Kristensen NP, Lees DC, Minet J, Mitter C, Mutanen M, Regier JC, Simonsen TJ, Wahlberg N, Yen S-H, Zahiri R, Adamski D, Baixeras J, Bartsch D, Bengtsson BÅ, Brown JW, Bucheli SR, Davis DR, De Prins J, De Prins W, Epstein ME, Gentili-Poole P, Gielis C, Hättenschwiler P, Hausmann A, Holloway JD, Kallies A, Karsholt O, Kawahara A, Koster JC, Kozlov M, Lafontaine JD, Lamas G, Landry J-F, Lee S, Nuss M, Park K-T, Penz C, Rota J, Schmidt BC, Schintlmeister A, Sohn JC, Solis MA, Tarmann GM, Warren AD, Weller S, Yakovlev RV, Zolotuhin VV, Zwick A (2011) Order Lepidoptera. In: Zhang Z-Q (Ed) Animal biodiversity: An outline of higher-level classification and survey of taxonomic richness. Zootaxa 3148: 212–221.
- Novotny V, Miller SE, Baje L, Balagawi S, Basset Y, Cizek L, Craft KJ, Dem F, Drew RAI, Hulcr J, Leps J, Lewis OT, Pokon R, Stewart AJA, Samuelson GA, Weiblen GD (2010) Guild-specific patterns of species richness and host specialization in plant-herbivore food webs from a tropical forest. Journal of Animal Ecology 79: 1193–1203. doi: 10.1111/j.1365-2656.2010.01728.x
- Peña C, Malm T (2012) VoSeq: A Voucher and DNA Sequence Web Application. PLoS ONE 7(6): e39071. doi: 10.1371/journal.pone.0039071
- Rambaut A, Drummond AJ (2007) Tracer v1.4. http://beast.bio.ed.ac.uk/Tracer
- Ratnasingham S, Hebert PDN (2007) BOLD: The Barcode of Life Data System (http://www.barcodinglife.org). Molecular Ecology Notes 7: 355–364. doi: 10.1111/j.1471-8286.2007.01678.x
- Ratnasingham S, Hebert PDN (2013) A DNA-Based Registry for All Animal Species: The Barcode Index Number (BIN) System. PLoS ONE 8(7): e66213. doi: 10.1371/journal. pone.0066213
- Ronquist F, Teslenko M, van der Mark P, Ayres DL, Darling A, Hohna S, Larget B, Liu L, Suchard MA, Huelsenbeck JP (2012) MrBayes 3.2: Efficient Bayesian Phylogenetic Inference and Model Choice Across a Large Model Space. Systematic Biology 61: 539–542. doi: 10.1093/sysbio/sys029
- Rota J (2005) Larval and pupal descriptions of the neotropical choreutid genera *Rhobonda* Walker and *Zodia* Heppner (Lepidoptera: Choreutidae). Annals of the Entomological Society of America 98: 37–47. doi: 10.1603/0013-8746(2005)098[0037:lapdot]2.0.co;2
- Rota J (2008a) Immature stages of metalmark moths from the genus *Brenthia* Clemens (Lepidoptera: Choreutidae): morphology and life history notes. Journal of the Lepidopterists Society 62: 121–129.

- Rota J (2008b) A new genus and new species of metalmark moths (Lepidoptera: Choreutidae) from Costa Rica. Zootaxa 1933: 12–18.
- Rota J (2011) Data partitioning in Bayesian analysis: molecular phylogenetics of metalmark moths (Lepidoptera: Choreutidae). Systematic Entomology 36: 317–329. doi: 10.1111/j.1365-3113.2010.00563.x
- Rota J, Wahlberg N (2012) Exploration of data partitioning in an eight-gene data set: phylogeny of metalmark moths (Lepidoptera, Choreutidae). Zoologica Scripta 41: 536–546. doi: 10.1111/j.1463-6409.2012.00551.x
- Stamatakis A, Hoover P, Rougemont J (2008) A rapid bootstrap algorithm for the RAxML web-servers. Systematic Biology 75: 758–771. doi: 10.1080/10635150802429642
- Taylor B, Maffi M (1978) A review of the mosquito fauna of the Solomon Islands (Diptera: Culicidae). Pacific Insects 19: 165–248.
- Wahlberg N, Wheat CW (2008) Genomic outposts serve the phylogenomic pioneers: designing novel nuclear markers for genomic DNA extractions of Lepidoptera. Systematic Biology 57: 231–242. doi: 10.1080/10635150802033006
- Wilson JJ (2012) DNA barcodes for insects. In: Kress WJ, Erickson DL (Eds) DNA Barcodes: Methods and Protocols. Springer, New York, 17–46. doi: 10.1007/978-1-61779-591-6\_3

## Appendix

Taxon descriptions organized in tabular format for ease of comparison.

Taxon	Niveas Rota, gen. n.	Niveas kone Rota, sp. n.	Niveas agassizi Rota, sp. n.
Type species	Niveas kone		
Material examined	See Table 1.	See Table 1.	See Table 1.
Material		The holotype and most paratypes will be retained at	The holotype will be deposited in National Museums
deposited		Agriculture Research Institute (Port Moresby), BMNH, Bishop Museum, Naturalis (Leiden), and	of Kenya (Nairobi) (NIMK), with paratypes to USNM, BMNH and NMK.
Distribution	Kenya, Papua New Guinea, Solomon Islands.	CSIRO (Canberra). Papua New Guinea, Solomon Islands.	Kenya.
Diagnosis	Niveas can be easily distinguished from most genera	N. kone can be separated from all other known	N. agassizi can be separated from other known
	of choreutids by the wing pattern (Figs 1, 3). Superficially, species of <i>Niveas</i> are similar to some	choreutids based on its wing pattern (Fig. 1). Superficially, it is similar to a few species of <i>Brenthia</i>	choreutids by the wing pattern (Fig. 3). It is superficially similar to some species of <i>Choreutis</i> .
	species of Anthophila and Choreutis, but there is	and <i>Litobrenthia</i> owing to its background color,	but the latter usually have forewings with apparent
	no known species in either of the latter two genera	but it lacks iridescent spots along forewing termen,	patterning, and this is absent in N. agassizi. Female
	with a black terminal band enclosing white spots	which are always present in those two genera. Both	genitalia are very distinct from those of other
	11, 3). Forewing as 111 1v. agassest and 1v. kone. (1.1gs	those of other choreutids (Figs 7, 8).	cnoredius (rig. 2).
	branches or with R <sub>4</sub> and R <sub>5</sub> fused in the basal half is		
	also diagnostic for the genus. Female genitalia with		
	paired concave sclerotizations on A7 sternite are also		
	unique to Niveas.		
Description	Figs 1–9.	Figs 1, 2, 5, 7, 8.	Male unknown. Figs 3, 4, 6, 9.
Head	Labial palpi with projecting ventral scale tufts (Figs	Fig. 2.	Fig. 4.
	2, 4).		
Wings	Forewing veins R four-branched in N. kone (Fig.	Fore- and hindwing with brown background color,	Forewing bronze-brown with speckled white-tipped
	5), five-branched in N. agassizi (Fig. 6), with R <sub>4</sub> an	speckled with white-tipped scales in an irregular	scales over most of its surface; distinct dark brown to
	R <sub>2</sub> fused in basal 3/5; CuP present at termen for	pattern; a distinct black band along termen of both	black band along termen with two small white spots
	1/3 to 1/5 wing length, extending as fold further	wings within which are more or less equidistant white	at apex; hindwing light brown (Fig. 3).
	towards base. Hindwing ten-veined, with M <sub>2</sub> in	spots (Fig. 1).	
	close proximity to the basally fused M <sub>3</sub> and CuA <sub>1</sub>		
	(N. agassizi) or nine-veined, apparently with M <sub>3</sub> and		
	CuA, completely fused into a single vein (Figs 5, 6).		

Taxon	Niveas Rota, gen. n.	Niveas kone Rota, sp. n.	Niveas agassizi Rota, sp. n.
Male genitalia	Tegumen rounded on top, tuba analis extending beyond tegumen; vinculum as inverted trapezoid ventrally emarginate; valva with costal margin straight, ventral margin rounded, ending with a horn-like projection; phallus twice as long as valva (Fig. 7).	As for the genus (Fig. 7).	Unknown.
Female genitalia	Apophyses anteriores slightly longer than posteriores; ostium bursae on A7 with a more or less strongly sclerotized antrum; ductus bursae straight, not coiled, with strong lateral sclerotizations; corpus bursae as a single sac (N. agassizi) or divided into two sacs (N. kone) with one or more signa. A7 sternite with paired, somewhat rounded, concave sclerotizations proximally, clearly visible in N. kone (Fig. 8), and slightly less so in N. agassizi (Fig. 9).	Corpus bursae split into two sacs; one sac with a V-shaped signum, the other with two round signa (Fig. 8).	Ductus bursae short and wide, opening into large corpus bursae, with one oval signum (Fig. 9).
Immature stages		Fig. 12. See a brief note in text.	Unknown.
Host plants	Genus Ficus (Moraceae).	Ficus botryocarpa Miq., F. nodosa Teijsm. & Binn., F. phaeosyce K. Schum. & Lauterb., F. pungens Reinw. ex Blume, F. variegata Blume, and F. wassa Roxb. (Moraceae).	Unknown.
Etymology	The generic name is derived from Latin <i>niveum</i> , meaning snowy, in reference to speckles of white-tipped scales in the wings of the type species; it is not treated as a Latin word and is feminine in gender.	The species is named after the Finnish Kone Foundation (Koneen Säätiö) in appreciation of their funding of this work. The name is a noun in apposition.	This species is named after David Agassiz, who collected all the known specimens and made many significant contributions to our knowledge of African micro-moths. The name is a noun in the genitive case.